

# ***Exposure to sub-10nm “particles” emitted from combustion sources: in-vitro toxicity and inflammatory potential***

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**20<sup>th</sup> ETH- Conference on Combustion Generated Nanoparticles**

13-16 June 2016

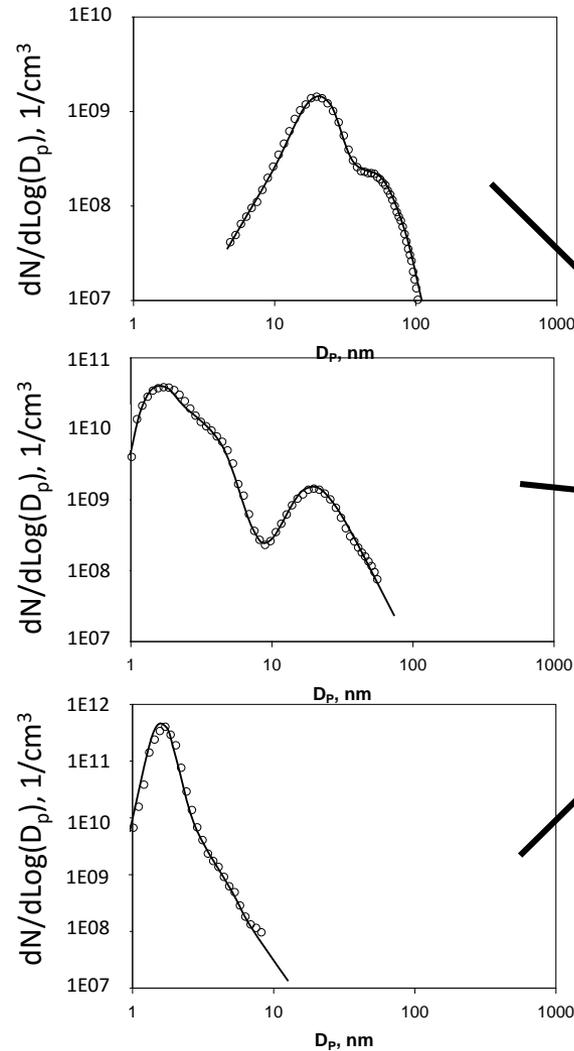


# Combustion Generated Nanoparticles

*premixed flames (ethylene/air - 1 bar)*

## DMA measurements

↑  
*residence time*

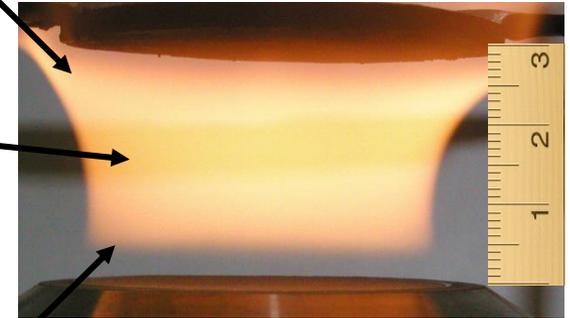
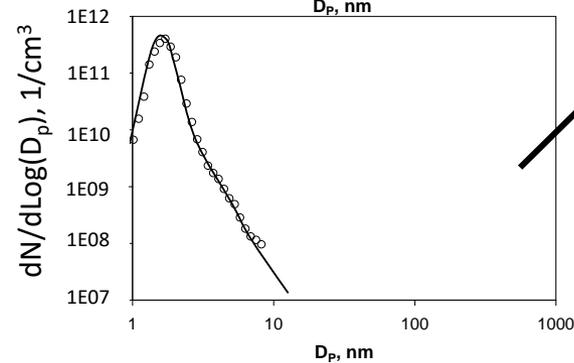
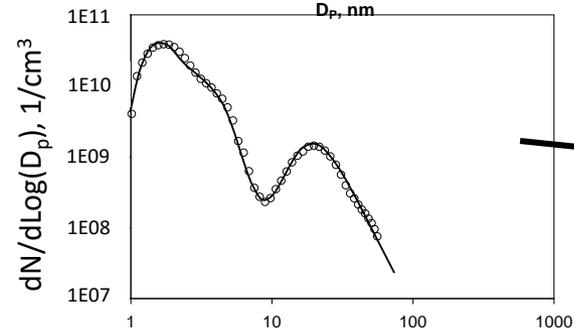
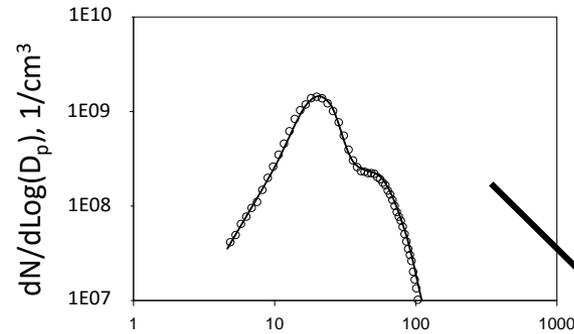
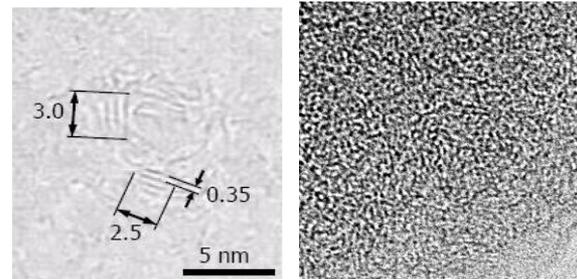
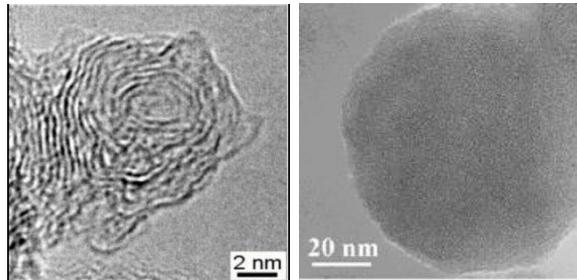
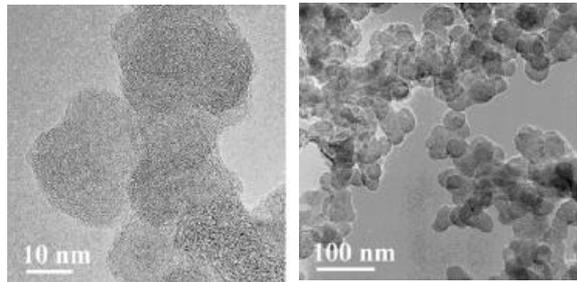


↑  
*residence time*

# Combustion Generated Nanoparticles

*premixed flames (ethylene/air - 1 bar)*

## Nano-DMA and HR-TEM

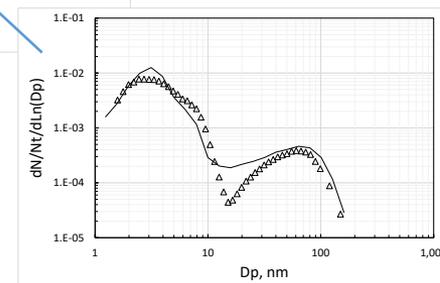
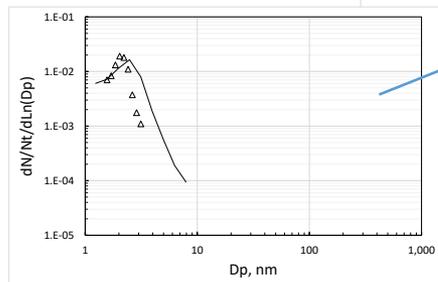
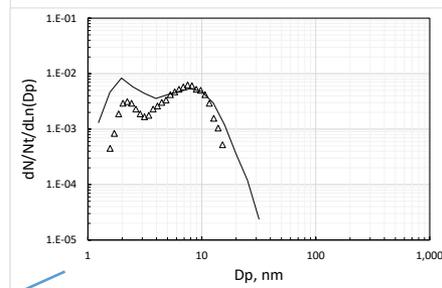
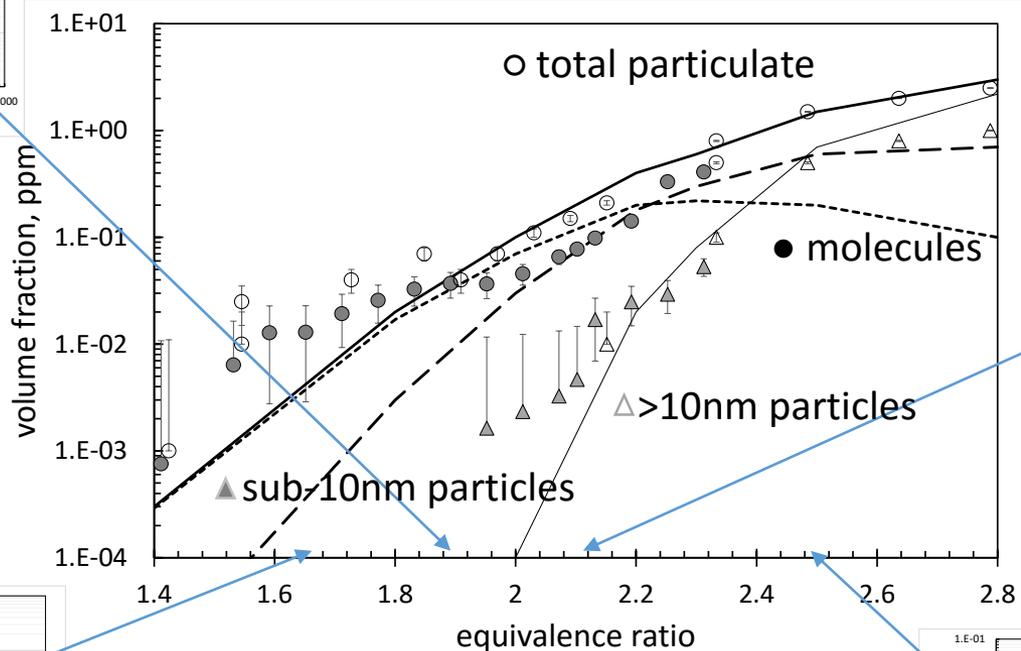
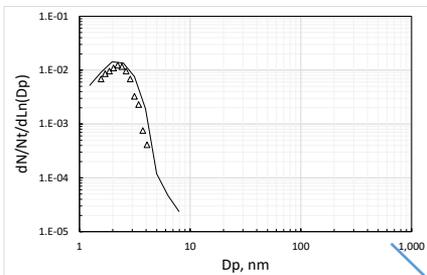


D'Anna, A. "Combustion-formed Nanoparticles", Proc. Combust. Inst. 32:593-613 (2009).

# Combustion Generated Nanoparticles

*premixed flames (ethylene/air - 1 bar)*

*Particle volume fractions vs equivalence ratio*

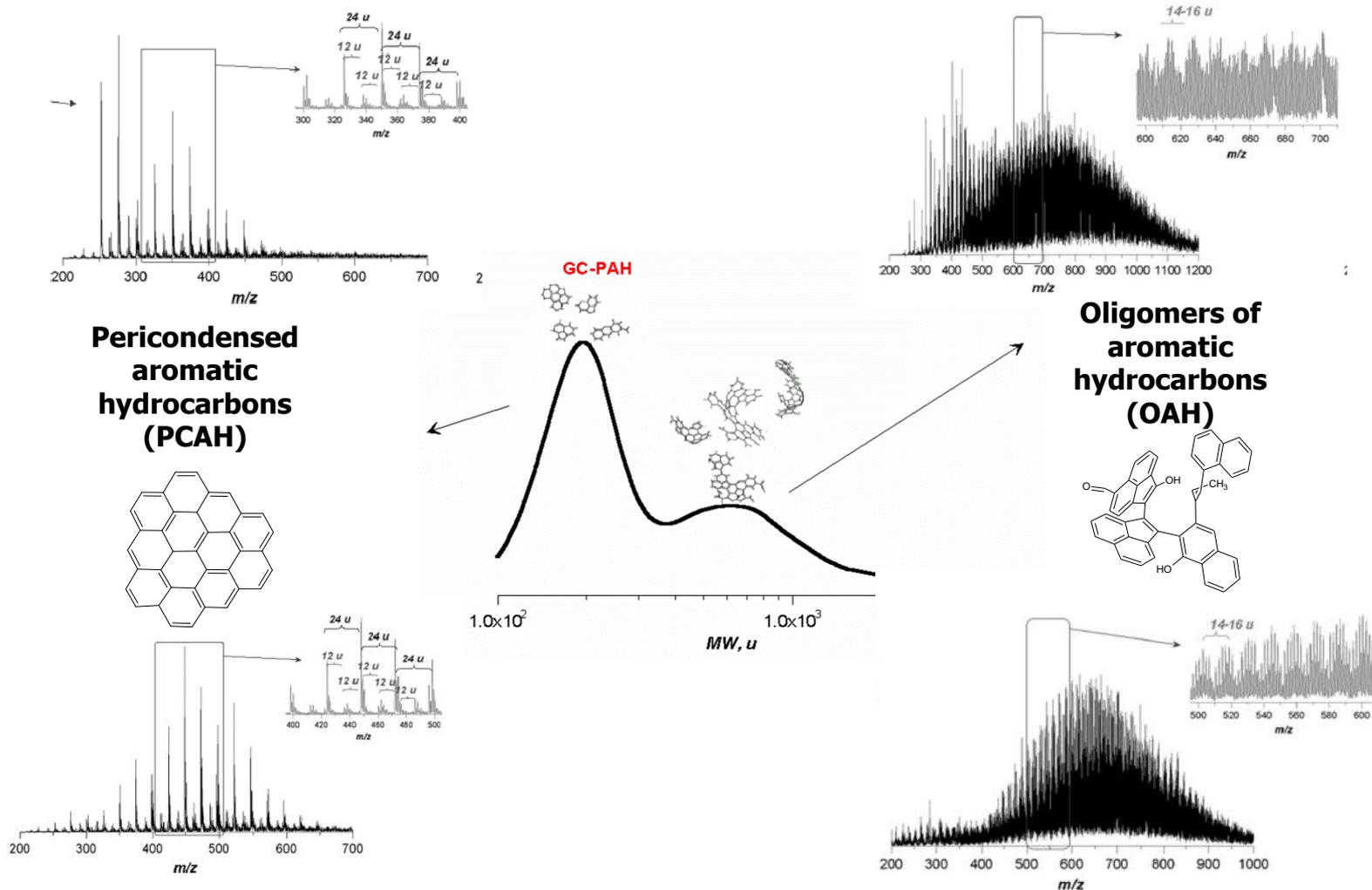


D'Anna A., "Kinetics of Soot Formation". 2015

in: Reedijk, J. (Ed.) Elsevier Reference Module in Chemistry, Molecular Sciences and Chemical Engineering. Waltham, MA: Elsevier.

# Sub-10 nm “particles”

MS – chemical characteristics

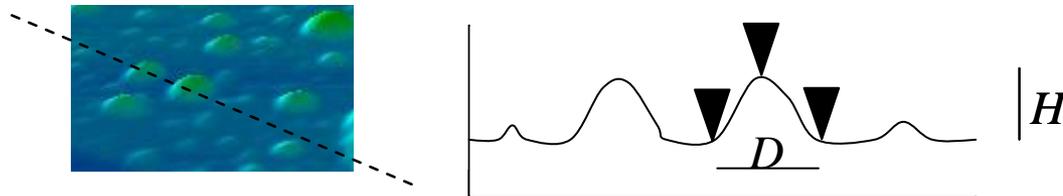
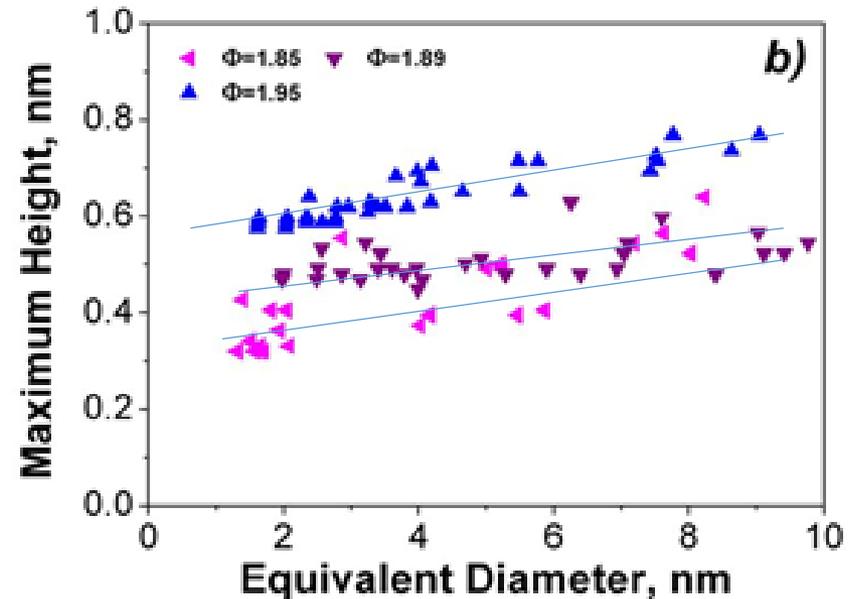
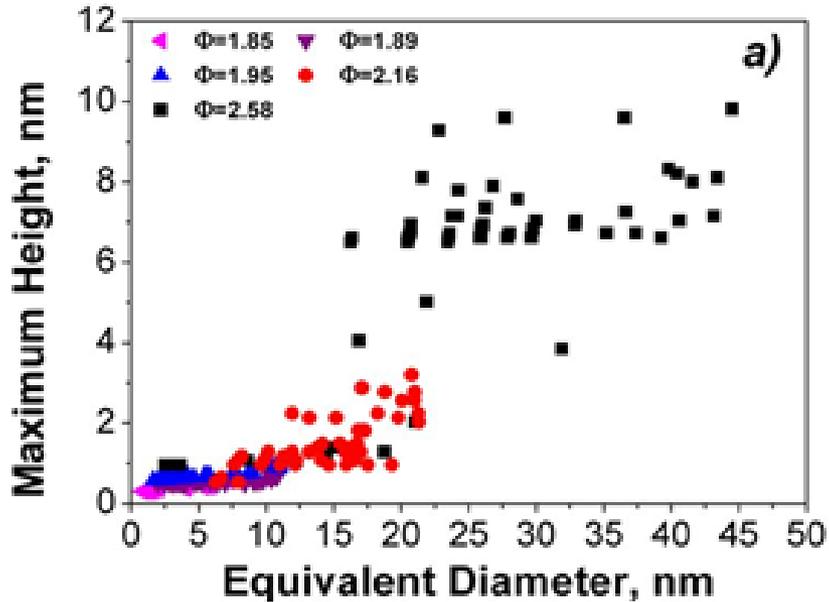


Ciajolo A., “Condensed phases in soot formation process”. (2009) Chapt. 21, 333-344

in Combustion Generated Fine Carbonaceous Particles, H. Bockhorn, A. D’Anna, A.F. Sarofim, H. Wang (Eds.), KIT Scientific Publishing,

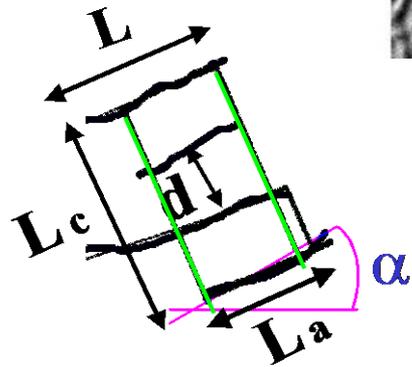
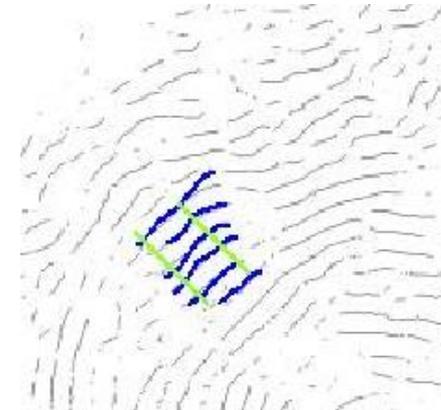
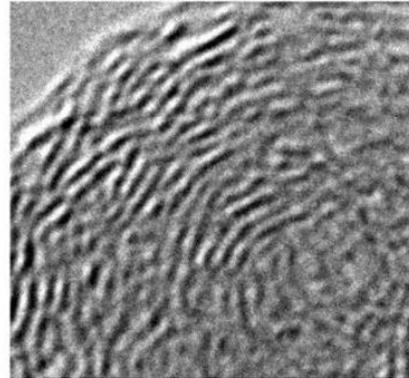
# Sub-10 nm “particles” morphology

## Atomic Force Microscopy



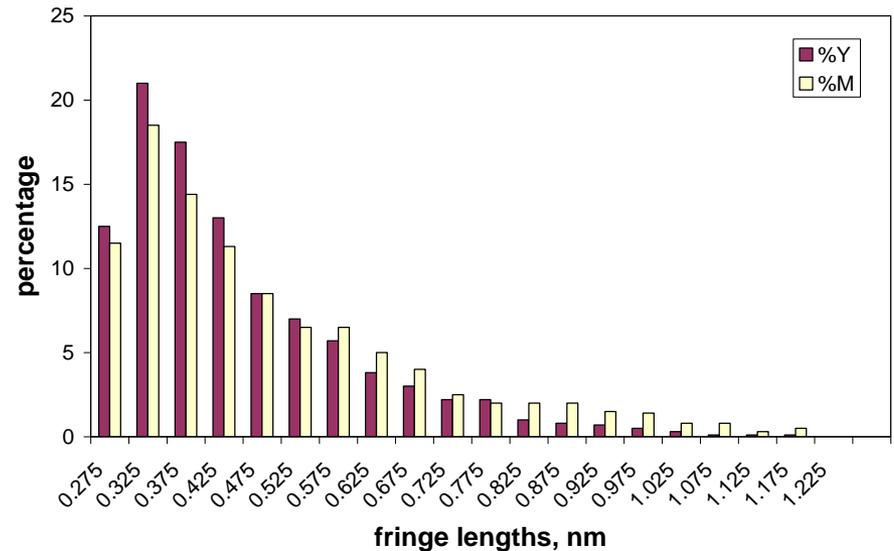
# >10 nm particles

## HRTEM – morphology (fringe analysis)

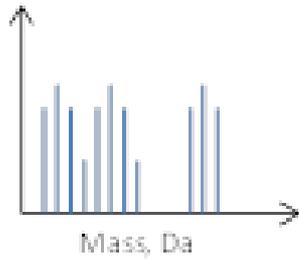


- L** mean fringe length
- La** diameter of the coherent domains (BSU)
- Lc** height of the BSU formed by **N** stacked layers with an interlayer spacing of **d**

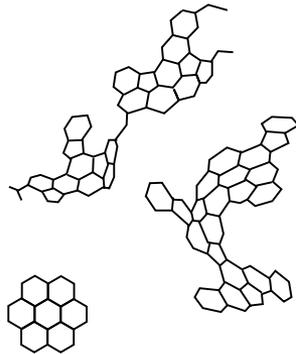
## skeletonization



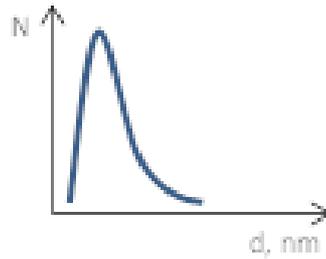
# Conceptual model for particle formation



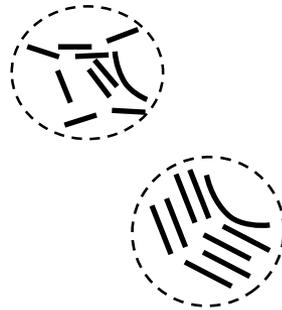
*nucleation*



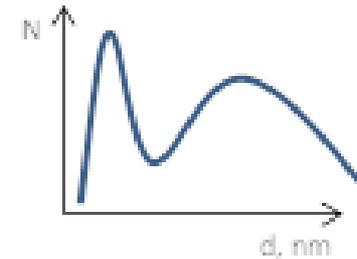
PAH molecules  
(200-1000 Da)



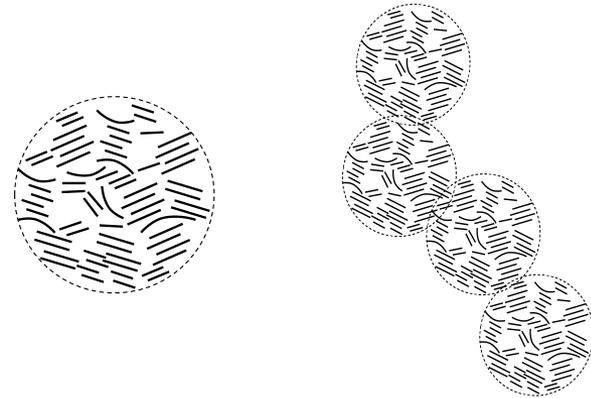
*coagulation  
(coalescence)*



cluster/nanoparticles  
(2-5 nm)



*aggregation*



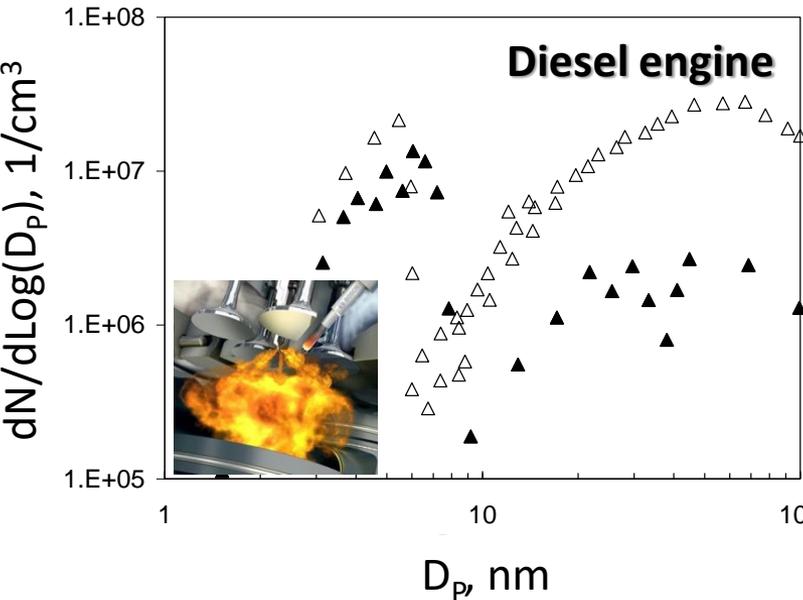
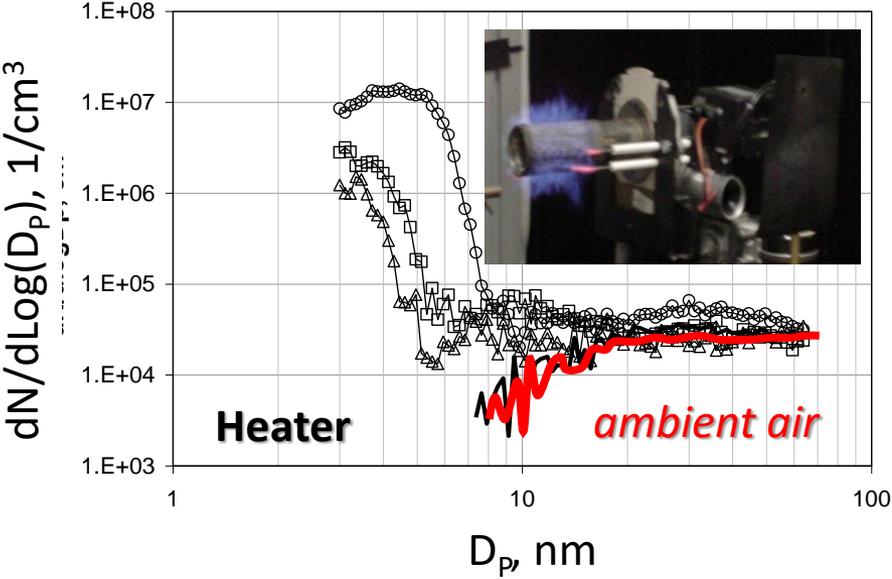
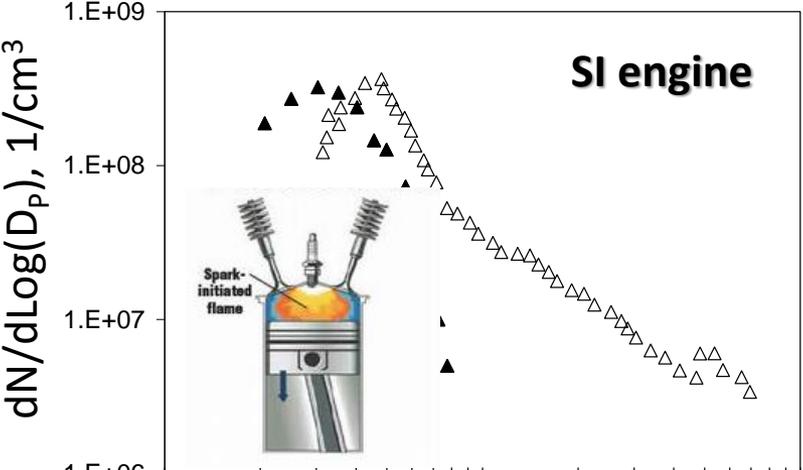
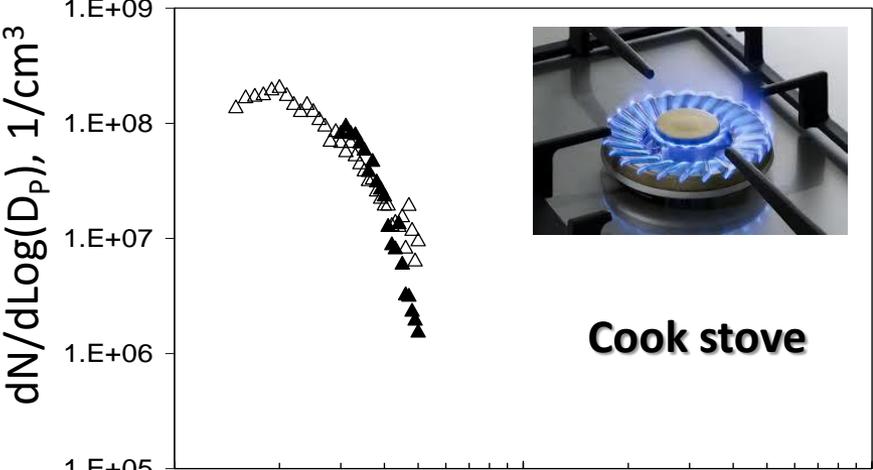
primary particles  
(15-30 nm)

aggregates  
(50-300 nm)

*sub-10nm*

*>10nm*

# Particle size distributions in practical systems



# Sampling and characterization procedures

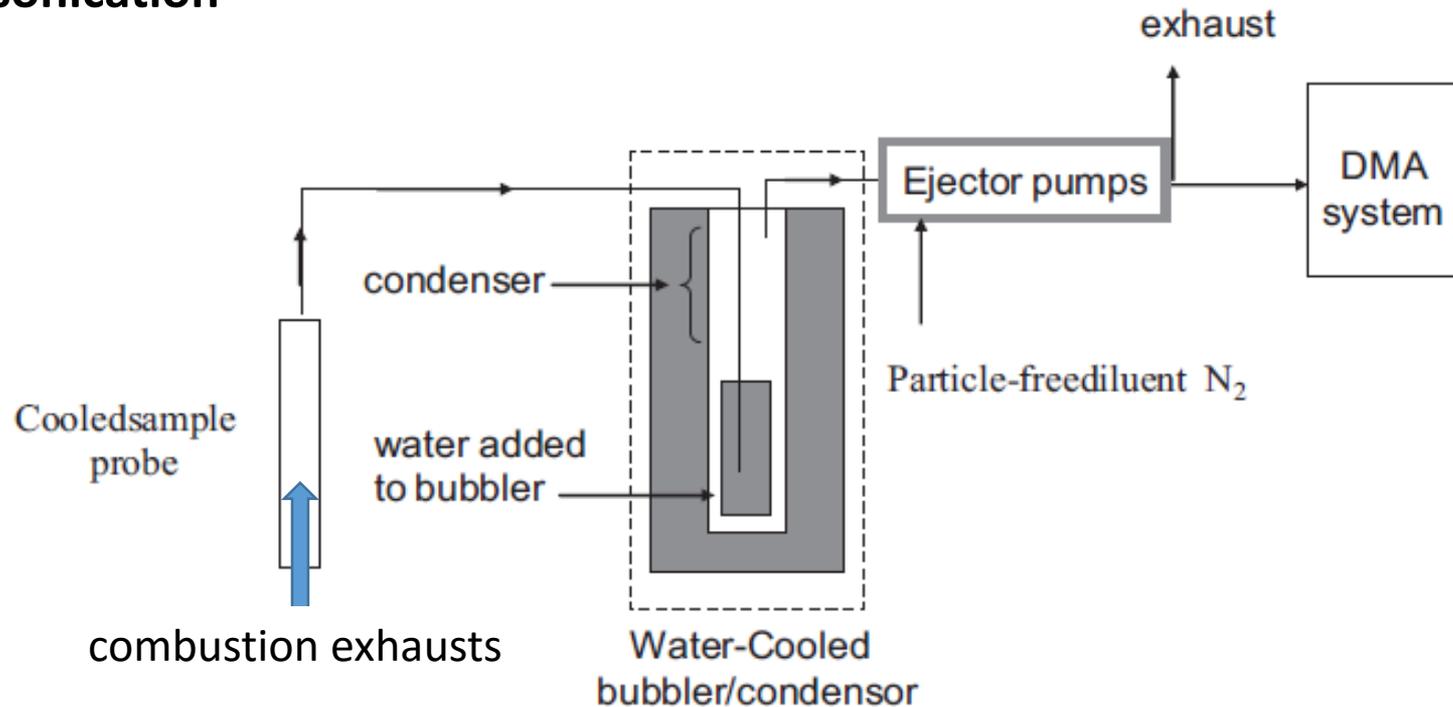
## sub-10nm “particles”

good water affinity

suspended in water with an efficiency of about 50% and isolated from the larger particles

separation of gaseous products by mild evaporation

ultrasonication



# Sampling and characterization procedures

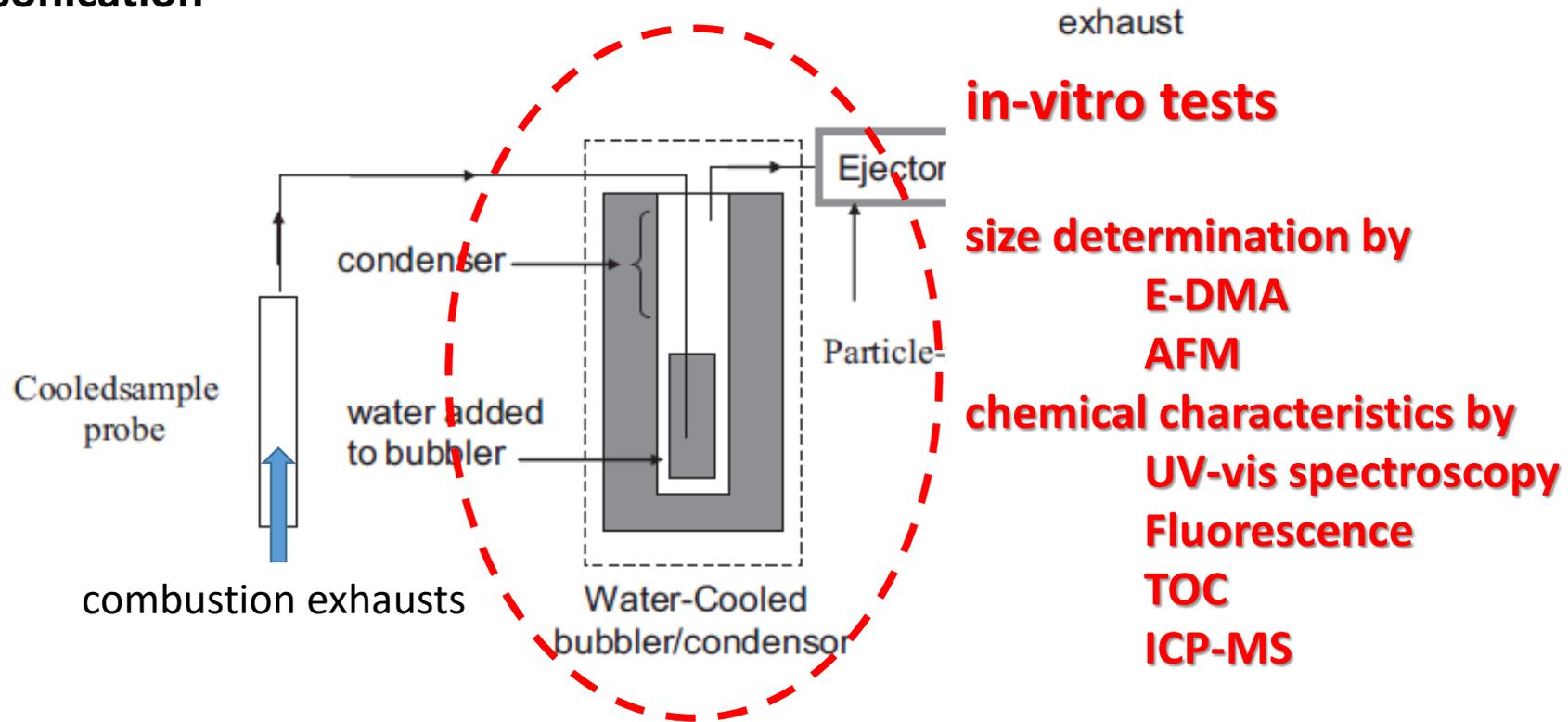
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# Sampling and characterization procedures

## >10nm particles

thermophoretic deposition on quartz plate

mechanical ablation of the solid material

stable suspension in water with 10% of DMSO

dispersion in solvent/solid matrix for structural characterization



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## >10nm particles

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## in-vitro tests

size determination by DLS

chemical characteristics by

UV-vis spectroscopy

FTIR

SEC

MS

# Cell culture

human alveolar epithelial-like cells (A549)

immortalized non-tumorigenic human dermal keratinocyte cells (HaCaT)

# Cell viability assay

cells were seeded in 96 well plate at a density of  $6 \times 10^3$  cells/well

culture media replaced with media containing sub-10nm particles at concentrations from 0.1 to 10 ppm

cells exposed to particles for 24 and 48 h

**cell viability evaluated by Cristal Violet Assay**

# Bioplex cytokine/chemokine detection

-cells were seeded in 6 well plate at a density of  $2 \times 10^5$  cells/well in DMEM 10%FCS

-culture media replaced with media containing sub-10nm particles at concentrations of 0.1, 1 ppm (blank test with DMEM)

-cells exposed to particles for 24 h

**pro- and anti-inflammatory cytokines released into the culture medium were detected (Bio-Plex Pro Human Cytokine 27-Plex Panel)**

## Apoptosi detection

-cells were treated with concentration of particles ranging from 0.1 to 10 ppm for 24 and 48 h

**Flow Cytometry - Annexin V-FITC apoptosis assay**

**Western-blot analysis of PARP1**

## Samples of sub-10 nm “particles”

	Cell viability	cytokine/ chemokine	flow cytometry & Annexin V- FITC/PI assay
Bluish, moderately-rich premixed flame	x		x
Cook-top burner	x	x	
Diesel engine exhaust (no DPF)	x	x	

## Samples of >10 nm particles

	Cell viability	Western-blot analysis of PARP1
Yellow, rich premixed flame	x	x

# Results

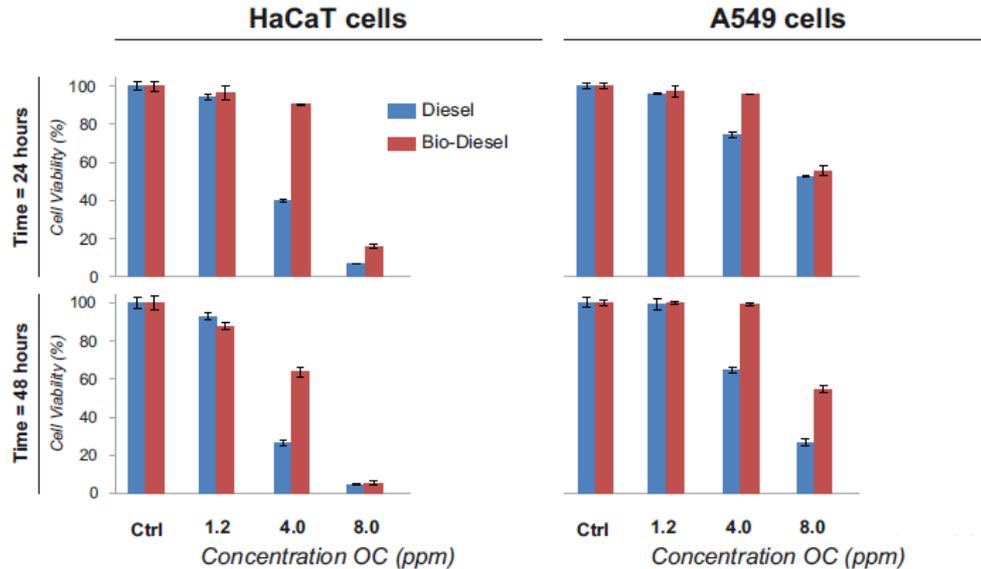
Cook-top burner **sub-10nm particles** show no relevant reduction in cell number on HaCat cells.

Results of secretome analysis suggest that the pro-inflammatory pathway is not activated for these nanoparticles.

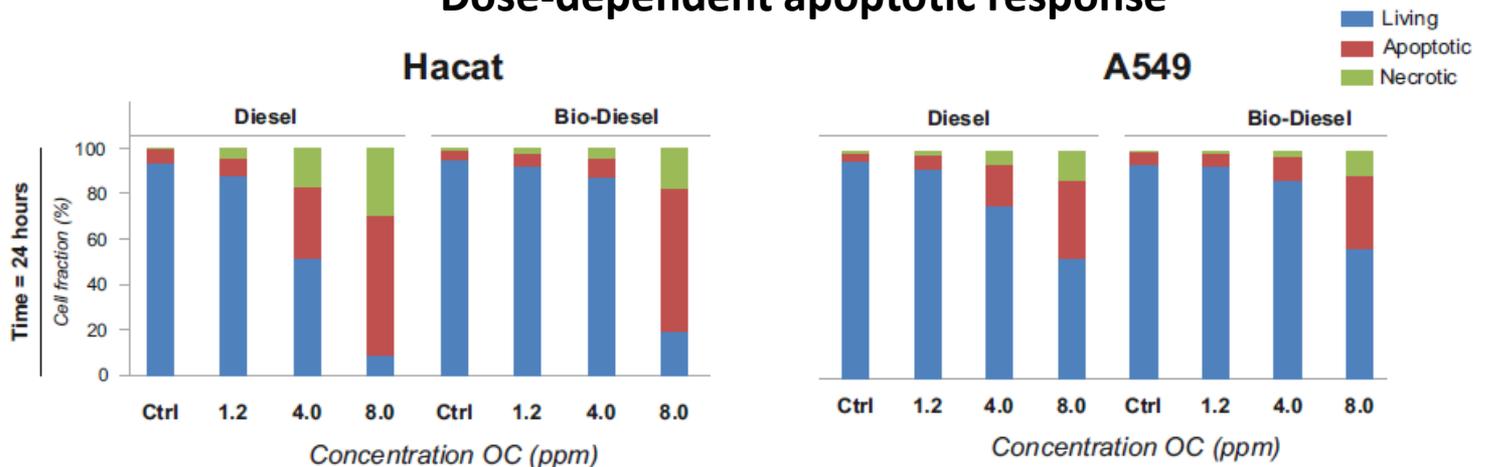
Flame generated **sub-10nm particles** appear to be significantly more dangerous. They also show to be more dangerous than engineered nanoparticles of the same sizes

# Results

Diesel and biodiesel generated **sub-10nm particles** induce cytotoxicity at lower concentrations with respect to flame formed particles.



## Dose-dependent apoptotic response



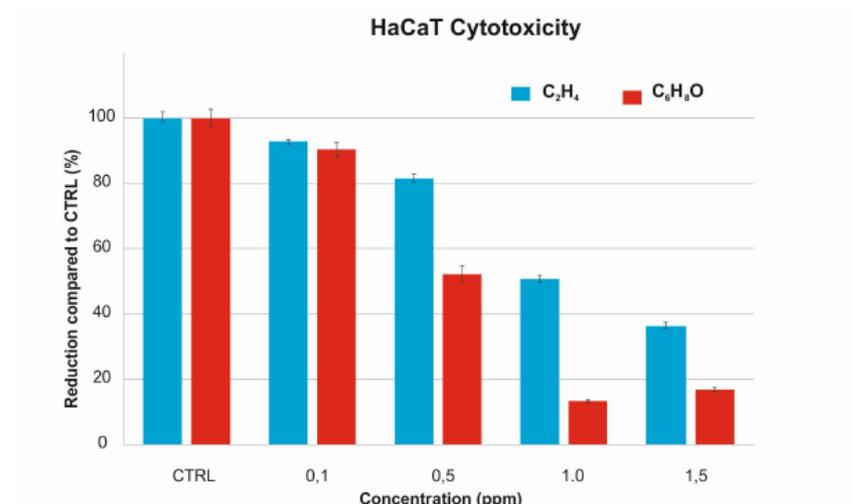
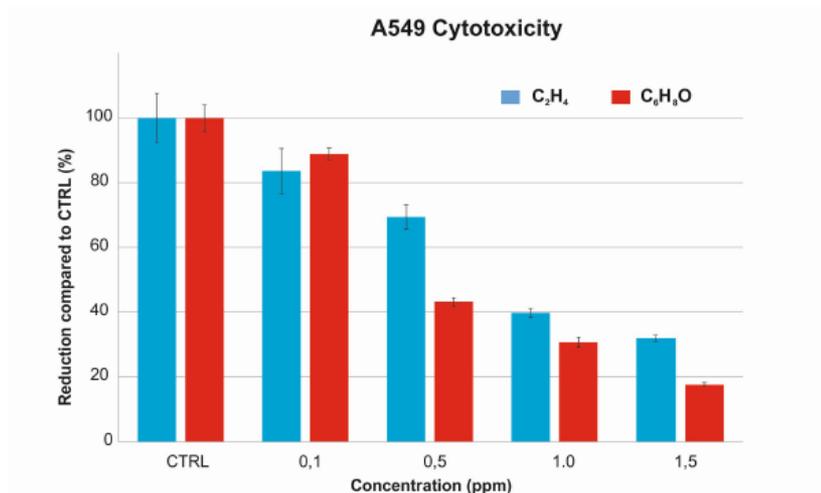
Diesel and Bio-Diesel fueled engine – **sub-10 nm particles**

# Results

Significant increase of cell mortality following treatment with **>10nm particles** for 24 hours in both cells lines

Dose-dependent effect

Biofuel **>10nm particles** seem induce more cytotoxicity compared to those emitted by the ethylene flame



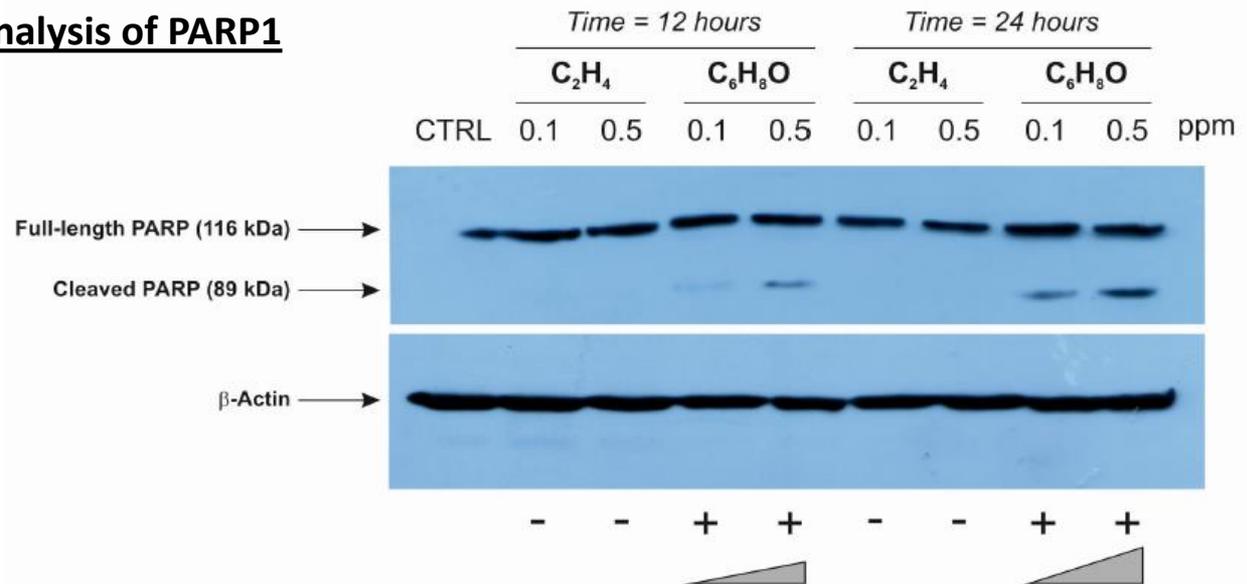
Flame generated particles in ethylene & ethylene/20% 2,5 dimethylfuran flames – **>10 nm particles**

# Results

Evidence of the presence of “p89” fragments of PARP1 in the cells exposed to **>10nm particles** generated by a ethylene/2,5 dimethylfuran flame

The **>10nm particles** derived from biofuel combustion have different chemical structure from the ethylene-generated particles: higher amount of oxygen atoms that substitute hydrogen atoms at the edges of the aromatic carbon network

## Western-blot analysis of PARP1



Flame generated particles in ethylene & ethylene/20% 2,5 dimethylfuran flames – **>10 nm particles**

# Conclusions

Combustion systems form and eventually emit huge number concentrations of **sub-10nm particles** which are clusters of aromatic compounds

Toxicity of these “particles” has been analyzed *in-vitro* with HaCaT and A549 cells

Toxicity of **sub-10nm particles** seems to be related to both particle size and particle chemical composition

(in the smaller particles the number of molecules at the surface of the cluster is comparable to the number of molecules constituting the cluster)

Ethylene/2,5dimethyl furan generated particles (**>10nm particles**) seem to induce stronger cytotoxicity than similar-size, ethylene generated particles (these particles have higher amount of oxygen atoms that substitute hydrogen atoms at the edges of the aromatic carbon network)